FOCAL ADHESION KINASE (FAK) INHIBITION AS A POTENTIAL STRATEGY FOR ANTICANCER THERAPIES

Y. Ma, B. van de Water, S. Le Dévédec*

Division of Toxicology, Leiden/Amsterdam Center Drug Research, Leiden University, Leiden, The Netherlands. *Correspondence: s.e.ledevedec@lacdr.leidenuniv.nl

CONTENTS

Abstract
Introduction
FAK and cancer479
Strategies for FAK inhibition in cancer therapy 479
Conclusions and prospective research48
References

ABSTRACT

Focal adhesion kinase (FAK) is a multidomain nonreceptor tyrosine kinase that mediates growth factor- and adhesion-derived cell signaling. FAK plays crucial roles in cell proliferation, survival, motility and invasion, all of which are hallmarks of cancer cells. Overexpression of FAK has been observed in diverse cancer types and is used as a marker for invasion and metastasis. Furthermore, in vivo animal studies demonstrated an involvement of FAK in tumor development and malignancy. Therefore, FAK is a potential target for anticancer drug discovery. In this review, we will first present FAK and its relationship to cancer, with focus on target validation of FAK. Secondly, approaches to inhibit FAK as a potential drug target for therapeutic intervention in cancer treatment will be discussed.

INTRODUCTION

Role of focal adhesion kinase

Cellular interactions with extracellular matrix and growth factors play essential roles in tumor initiation, progression and metastasis. Focal adhesion kinase (FAK) is a 125-kDa multidomain nonreceptor protein-tyrosine kinase (PTK) that mainly localizes in the cytoplasm of cells. Upon cell adhesion on diverse extracellular matrices and/or activation by growth factors, FAK is recruited to focal adhesions (FAs), the closest contacts between the cell and the extracellular matrix, and mediates FA signaling. The complex structure of FAK results in a broad range of protein–protein interactions with other tyrosine kinases, cytoskeletal and adaptor proteins that are part of the so-called adhesome (1). FAK is known to play important roles in

tumor progression and metastasis through its regulation of cancer cell migration, invasion, anchorage-dependent cell proliferation and survival. Recently, numerous in vivo studies have demonstrated the role of FAK in tumor initiation, as well as progression. In agreement with these experimental data, FAK is linked to human cancer mainly due to its overexpression and activation in a number of human tumors. Altogether these studies suggest that FAK is a potential target for drug discovery.

FAK structure and its regulation

As shown in Figure 1, FAK consists of several domains. The FERM domain (band 4.1-Ezrin-Radixin-Moesin homology) at the N-terminus negatively regulates the catalytic activity of FAK (2). FERM interacts with integrins and growth factor receptors, and through this domain FAK also binds the Arp2/3 complex to control actin assembly (3). In the catalytic kinase domain, autophosphorylation at the FAK Tyr397 residue recruits Src at the FA site. Furthermore, Src phosphorylates FAK at Tyr576 and Tyr577, which results in a conformational change that enhances the catalytic kinase activity. pTyr861 increases the binding affinity of p130cas to the proline-rich regions (PRRs) in the FAK C-terminus and is crucial to sense mechanical force (4) and H-Ras-induced transformation (5). The FA-targeting (FAT) domain at the C-terminal region is responsible for FAK localization to FA (6) and spatially interacts with paxillin and talin. FAK PRR binds Src homology 3 (SH3) domain-containing proteins such as p130cas, the GTPase regulator associated with FAK (GRAF) and the ARF GTPase-activating protein ASAP1. Phosphorylation of the Tyr925 residue in the FAT domain promotes GRB2 binding to FAK, which activates the mitogen-activated protein kinase (MAPK) pathway through FAK-GRB2-Ras-MEK1-ERK2. pTyr925 is also responsible for the cell survival function of FAK and mediates an MAPK-associated angiogenic switch during tumor progression (7, 8).

Recently, it was reported that pTyr407 negatively regulated kinase activity and cell migration/invasion (9, 10). Most of the tyrosine phosphorylated residues of FAK have been well studied, while the role of serine phosphorylated residues (Ser722, Ser846, Ser910) is still not very well understood (11, 12). Recently, it has been shown that phosphorylation at Ser732 in endothelial cells plays a role in the regulation of the centrosome during mitosis and this may contribute

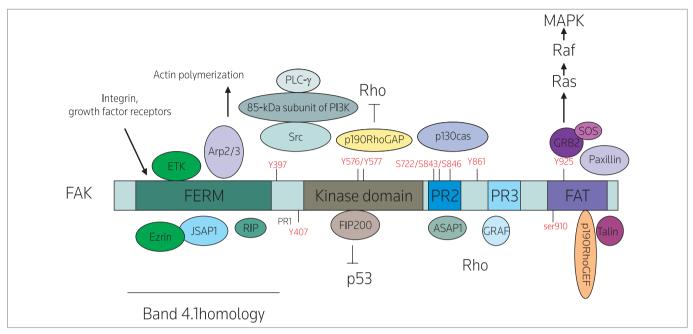


Figure 1. Focal adhesion kinase (FAK) structural features and binding partners. The kinase domain of FAK is flanked by the *N*-terminus that harbors the FERM domain and by the *C*-terminus that consists, in addition to proline-rich (PR) domains, of the FAT domain. The *N*-terminal domain has the Y397 autophosphorylation site, which is also the target site for different small-molecule inhibitors. The kinase domain has the Y576/577 tyrosines, important for the catalytic activity of FAK. The *C*-terminal part of FAK has Y861 and Y925 tyrosines. Different proteins bind to these domains and are involved in cell proliferation, motility and survival signaling.

to cell proliferation and angiogenesis (13). Ser843 is known to be phosphorylated when FAs disassemble and cells detach from the substratum, probably via inhibition of pTyr397 (14).

FAK in cellular processes

Processes related to tumor formation

FAK contributes to tumorigenesis through the promotion of cell survival and/or proliferation, which are hallmarks of cancer cells. FAK mediates survival signaling, for instance, through the PKB prosurvival pathway. FAK phosphorylation (pTyr397) is involved in doxorubicin-induced cell apoptosis in a Bcl-2- and caspase-independent manner. Indeed, inducible FAK-related non-kinase (FRNK) expression sensitizes cells to doxorubicin-induced apoptosis and inhibits doxorubicin-induced PKB activation (15). FAK also regulates cell proliferation by upregulation of some cyclins downstream of PKC, phosphatidylinositol 3-kinase (PI3K)/Akt and MAPK/ERK pathways (16-18). The FERM domain controls FAK function (19) and deletion of the FERM domain increases FAK phosphorylation and activity and affects cell cycle progression in CHO cells (2).

In addition to its role in FAs, FAK also plays a scaffolding role in the nucleus of cells under cellular stress conditions. FAK facilitates p53 degradation by its FERM domain-mediated nuclear localization and promotes cell proliferation and survival (19-21).

Processes related to tumor progression and metastasis

FAK controls FA assembly/disassembly at the leading edge of lamellipodia and disassembly at the rear of migrating cells. The role of FAK in cell motility is well documented (see reviews 6, 12, 22, 23). FAK interacts not only with integrins, but also with growth factor receptors. EGFR (erbB-1) and erbB-2 receptor signaling, which are also prognostic markers for cancer progression, regulate FA turnover, cell migration and invasion through the Src-FAK pathway (24, 25). Cellular traction forces cause dynamic conformational changes in the FERM domain that are shown to be involved in cell spreading and motility (26). A single residue, Lys38, at the subdomain F1 is important for the kinaseinhibitory effect of FERM and the mutant K38A increases FAK phosphorylation, cell cycle entry and cell migration (27). The F2 subdomain of the FAK FERM domain and phosphorylation of Tyr1349 at c-Met are critical for c-Met binding, and this interaction fully activates FAK activity and HGF-induced c-Met-mediated cell invasion (28). FRNK lacks kinase activity and has a competitive inhibitory effect on FAK at focal contacts. FRNK expression abolishes EGF- or v-Src-induced activation of downstream molecules (ERK, JNK, etc.) and cell motility, and prevents cell invasion by inhibiting matrix metalloproteinase (MMP) secretion (29, 30). Inducible ectopic expression of FRNK sensitizes the human embryonic kidney (HEK) cell line to 5-fluorouracil and decreases haptotactic mobility (31). Also, disruption of FAK by FRNK decreases cell attachment, motility and invasion in head and neck squamous cell carcinoma by downregulation of MMP-2 expression (32).

Introduction of the FAT domain, which also competes for binding of endogenous FAK at FA sites, has been reported to sufficiently inhibit cell invasion and sensitize cells to apoptotic stimuli (33). The aggressive phenotype of prostate cancer cells depends on FAK expression, which is regulated by ERK signaling (34). In addition, FAK is also involved in tumor invasion and angiogenesis by regulation of MMP and vascular endothelial growth factor (VEGF) expression (8, 29).

FAK AND CANCER

Increased FAK expression in human cancers

In agreement with the experimental data, overexpression of FAK has been reported in a wide range of cancers (reviewed in 20, 35-37), such as breast (38-42), head and neck, cervical (38), colon (43, 44), thyroid (45), prostate (46), liver (47, 48), skin, lung, bone, melanoma (49) and ovarian cancers (50). FAK apparently plays distinct roles depending on the tumor type and development stage. Similar observations have been obtained in cell lines derived from tumor and normal tissue (51, 52). These studies show that FAK overexpression plays a role in tumor formation and metastasis and could be used as a prognostic marker (38, 42, 45, 53, 54).

Role of FAK in tumor development

Many recent studies using both mouse xenograft models and conditional knockout (KO) mice have provided evidence that FAK signaling pathways can stimulate tumor initiation, as well as tumor progression and metastasis, through their regulation of cell migration, invasion, epithelial-to-mesenchymal transition and angiogenesis.

FAK in promoting tumor initiation

FAK KO animals die, and to overcome embryonic lethality conditional tissue-specific FAK deletion in vivo has been developed with Cre recombinase (Cre)/loxP strategies, including myosin light chain 2a(MLC2v)-Cre/FAK^{flox} (55), Cre-ER (estrogen receptor)/FAK^{flox} (15, 56, 57) and MLC2a mouse mammary tumor virus (MMTV)-Cre/FAKflox (58). The first experimental proof using conditional FAK deletion in the epidermis demonstrated the role of FAK in skin tumorigenesis (56). The intercross of FAKflox/flox mice with MMTV-Cre mice resulted in the deletion of FAK in mammary epithelial cells, which disturbed mammary gland development (59). Complete deletion of FAK in both luminal and myoepithelial mammary epithelial cells disrupted mammary gland formation (van Miltenburg, In press). Using the mouse polyomavirus middle T (PyVmT) transgenic breast cancer model, conditional floxed FAK was introduced. In the MMTV-Cre/PyVmT mice, the mammary epithelium-specific disruption of FAK altered the FAK-related signal cascades and retarded tumor initiation and progression, as well as lung metastasis formation (60). FAK deletion also reduced the population of mammary cancer stem/progenitor cells (61) and disrupted the transition of premalignant hyperplasias to carcinomas and subsequent metastasis (58).

200 kDa FAK family kinase-interacting protein (FIP200), also known as RB1-inducible coiled-coil 1, was identified as an inhibitor of FAK and Pyk2 by interaction with their kinase domains (62, 63). FIP200 inhibits FAK/Pyk2-related cellular functions and interacts with RB1 and p53 (64). Large truncation deletion of the *RB1CC1* gene has been observed in 20% of primary breast cancers and this indicates a possible role for FIP200 as a tumor suppressor (65).

FAK in promoting tumor progression and metastasis

Several studies in mouse xenograft models provide evidence that FAK is required for both tumor progression and metastasis. Overexpression of FAK in human malignant astrocytoma cells contributed to an increase in tumor volume due to enhanced cell proliferation (66). After

tail vein injection of v-Src-transformed NIH/3T3 fibroblasts (30) or B16F10 mouse melanoma cells (67), stable expression of FRNK results in a reduction of experimental lung metastases. Recently, our group demonstrated that FRNK sensitizes rat breast cancer MTLn3 cells to doxorubicin in vitro and in vivo via inhibition of doxorubicin-induced PKB activation and downregulation of FRA-1 (a member of the activator protein-1 complex) (15). Inducible expression of FRNK inhibits cell spreading and migration in vitro, as well as primary tumor formation and the early phase of metastasis, but not the outgrowth of macrometastases (68). In agreement with these results, by using FAKnull or knockdown cells, others have confirmed that the catalytic activity of FAK is required for metastatic breast cancer progression (8, 69, 70) and that FAK signaling is critical for erbB-2/erbB-3 receptor cooperation for oncogenic transformation and invasion (71) and for Ras- and PI3K-dependent breast tumorigenesis in mice (72). Pharmacological inhibition of FAK in wild-type mice suppressed angiogenesis, which is a key element in tumor progression and metastasis (57).

STRATEGIES FOR FAK INHIBITION IN CANCER THERAPY

In vitro and in vivo studies, as well as clinical studies, support the fact that FAK is a potential target for the treatment of cancer. Currently, inhibition of FAK signaling is under investigation for its beneficial effects in cancer treatment. FAK signaling can be disturbed by affecting, in practice, either FAK expression or FAK activation.

Agents inhibiting FAK expression

Interfering RNAs

FAK short interfering RNA (siRNA) shows high efficacy in gene expression downregulation in in vitro studies (73, 74). Inhibition of FAK with short hairpin RNAs (shRNAs) prevents FAK function in cell adhesion, migration and proliferation in the highly invasive human prostate cancer cell line PC-3M and the mouse breast cancer cell line 4T1 in vitro, and suppresses tumor growth in heterotopic/orthotopic mouse models in vivo (75). Moreover, knockdown of FAK and the family member Pyk2 extended survival in orthotopic glioma xenograft models in mice (76). However, because of its fast degradation, it is difficult to deliver siRNA very efficiently in vivo, questioning the clinical practicability of using siRNA. A modified polyethylenimine (PEI) gene carrier can be used to deliver FAK shRNA in vitro and in vivo to study the effect of FAK inhibition as a melanoma therapeutic (77). Recently, a neutral lipid liposome, 1,2-dioleoyl-sn-glycero-3-phosphatidylcholine (DOPC), has been reported to introduce FAK siRNA successfully and silencing of FAK showed antitumor and antiangiogenic effects; specifically, it improved the chemosensitivity to docetaxel or cisplatin in an orthotopic ovarian carcinoma model (78). This type of lipid liposome improved intratumoral penetration, delivery efficacy and toxicity. If the efficiency of specific RNA delivery can be improved, the strategy of inhibiting FAK expression is promising for cancer treatment.

(In)direct pharmacological modulation of FAK activity

Receptor tyrosine kinase inhibitors

FAK phosphorylates downstream effectors, and the use of kinase inhibitors consequently disturbs signaling transduction. The EGFR

inhibitor gefitinib (Iressa®) and the PTK inhibitor genistein are already used in the clinic. Gefitinib was shown to decrease FAK phosphorylation and inhibit the metastasis of oral squamous cancer cells to the lymph nodes (79). Genistein increased the adhesion of prostate cancer cells by modulating FAK activity (80). In addition, herbimycin A, another PTK inhibitor, decreased FAK phosphorylation and resulted in decreased migration of oral squamous cancer cells (81). Trastuzumab (Herceptin®), which binds to the juxtamembrane region of the erbB-2 receptor, was shown to inhibit Src and FAK activities, thereby inhibiting FA turnover, which was evident by increased FA stability and reduced cell invasion (24). Bosutinib (SKI-606), a novel Src kinase inhibitor, inhibits the phosphorylation of FAK and suppresses the migration and invasion of human breast cancer cells (82). Dasatinib (BMS-354825) inhibits the migration and invasion of non-small cell lung cancer and head and neck squamous cell carcinoma cells. The effects on migration and invasion correlated with the inhibition of Src and downstream mediators of adhesion such as FAK, p130cas and paxillin (83).

Small-molecule inhibitors

Several FAK inhibitors have been developed by pharmaceutical companies (see Table I), the most promising of which are PF-562271 (Pfizer) and TAE-226 (Novartis) (26). These are ATP analogues and effectively inhibit the kinase activity of FAK. Treatment of cells with these inhibitors results in a decrease in Tyr397 phosphorylation, which is associated with inhibition of cell migration (84, 85).

PF-562271 has a dual effect on FAK and Pyk2, with a nanomolar IC_{50} in vitro (86). In vivo maximal inhibition of pFAK (78%) is obtained 1 h after a dose of 33 mg/kg p.o. in tumor-bearing mice. After a single dose, > 50% inhibition of FAK phosphorylation lasts for over 4 h. Furthermore, dose-dependent tumor growth inhibition and regression were observed in a broad range of human s.c. xenograft models, including prostate, breast, pancreatic, colon,

lung and glioblastoma tumors, with no observation of weight loss, morbidity or death.

The in vivo inhibitory mechanisms of PF-562271 rely on anoikis/apoptosis and reduction of microvascular density (86). Recently, in a rat bone model of human cancer, this compound was shown to inhibit breast cancer MDA-MB-231 growth in the bone without altering normal bone formation (87). Initial phase I data on PF-562271 from patients with different types of cancer revealed that the compound appears to be performing well (88).

TAE-226 is another dual inhibitor specific for both FAK and insulinlike growth factor 1 receptor (IGF-I receptor), so a direct effect due to FAK inhibition cannot be determined. Nevertheless, it is a novel bisanilinopyrimidine inhibitor that is reported to efficiently inhibit FAK signaling, arrest tumor growth and invasion, and prolong the lifespan of mice with glioma or ovarian tumor implants (84, 89, 90). TAE-226 induces an intermediate conformation of the kinase activation loop (91), which inhibits the phosphorylation of FAK and the downstream molecules Akt and ERK. TAE-226 also decreases cell proliferation, adhesion, migration and invasion in glioma cells (84). It has significant activity in ovarian carcinoma and inhibits pTyr397, pTyr861 and cell growth in a time- and concentration-dependent manner. Moreover, in tumor-bearing mice it shows a synergistic effect with docetaxel on reduction in cell growth and tumor burden and increase in survival (89). TAE-226 is also promising in the therapy of imatinib-resistant gastrointestinal stromal tumors (92). A phase I clinical trial study is under way.

In November 2008, Pfizer initiated a phase I clinical study with another inhibitor, PF-04554878, in patients with advanced nonhematological malignancies.

The primary mechanisms of action of FAK inhibitors in vivo are apoptosis/anoikis, antiangiogenesis and reduced invasion. However, the role of specific FAK inhibitors in vivo and their possible long-term side effects should be investigated.

Table I. FAK inhibitors in cancer research.

Inhibitor	In vitro/in vivo effects	Ref.
PF-562271* (FAK/Pyk2)	Xenograft models and patients (antiangiogenic, antitumor activity) Phase I clinical trial (Pfizer)	(86, 88) http://clinicaltrials.gov/ct2/show/NCT00666926
TAE-226* (FAK/IGF-I receptor)	Breast cancer, glioblastoma, gastrointestinal stromal tumors, ovarian carcinoma, esophageal adenocarcinoma (cell proliferation/growth, cell cycle, cell migration, chemoresistance to docetaxel in vitro and in vivo) Phase I clinical trial (Novartis)	(78, 84, 88, 90, 92, 98)
PF-573228* (FAK)	Human tumor cell lines (cell migration)	(85, 96)
TAC-544* (FAK/Pyk2)	Animal study (angiogenesis)	(58)
1,2,4,5- Benzenetetraamine tetrahydrochloride* (FAK)	Breast cancer (cell adhesion, tumor regression)	(99)
PF-04554878# (FAK/Pyk2)	Nonhematological malignancies Phase I clinical trial (Pfizer)	http://clinicaltrials.gov/ct2/show/NCT00787033

^{*}ATP analogues that inhibit FAK kinase activity; #unknown inhibitory mechanism.

CONCLUSIONS AND PROSPECTIVE RESEARCH

FAK, a central protein in FA sites, regulates various cellular processes, including cell proliferation, survival, migration and invasion, which are crucial steps in tumorigenesis and metastasis formation. The implications of FAK overexpression in cancer suggest that FAK inhibition is a potential target for anticancer therapy. More and more evidence shows that FAK inhibition alone and in combination with other traditional therapies is a promising strategy in cancer treatment (74, 93-97). This review summarizes the latest studies and findings on genetic, functional and pharmaceutical inhibition of FAK. Future work should focus on the development and clinical evaluation of new inhibitors, as well as modulation of target-specific delivery.

DISCLOSURE

We thank all members in the Division of Toxicology for their helpful discussions. This work was financially supported by grants from the Dutch Cancer Society (UL 2006-3538 and UL 2007-3860).

REFERENCES

- 1. Zaidel-Bar, R., Itzkovitz, S., Ma'ayan, A., Iyengar, R., Geiger, B. Functional atlas of the integrin adhesome. Nat Cell Biol 2007, 9(8): 858-67.
- 2. Cooper, L.A., Shen, T.-L., Guan, J.-L. *Regulation of focal adhesion kinase* by its amino-terminal domain through an autoinhibitory interaction. Mol Cell Biol 2003, 23(22): 8030-41.
- 3. Serrels, B., Serrels, A., Brunton, V.G. et al. Focal adhesion kinase controls actin assembly via a FERM-mediated interaction with the Arp2/3 complex. Nat Cell Biol 2007, 9(9): 1046-56.
- 4. Sawada, Y., Tamada, M., Dubin-Thaler, B.J. et al. Force sensing by mechanical extension of the Src family kinase substrate p130Cas. Cell 2006, 127(5): 1015-26.
- Lim, Y., Han, I., Jeon, J., Park, H., Bahk, Y.-Y., Oh, E.-S. Phosphorylation of focal adhesion kinase at tyrosine 861 is crucial for Ras transformation of fibroblasts. J Biol Chem 2004, 279(28): 29060-5.
- 6. Mitra, S.K., Hanson, D.A., Schlaepfer, D.D. Focal adhesion kinase: In command and control of cell motility. Nat Rev Mol Cell Biol 2005, 6(1): 56-68.
- 7. Arold, S.T., Hoellerer, M.K., Noble, M.E.M. *The structural basis of localization and signaling by the focal adhesion targeting domain.* Structure 2002, 10(3): 319-27.
- 8. Mitra, S.K., Mikolon, D., Molina, J.E. et al. *Intrinsic FAK activity and Y925 phosphorylation facilitate an angiogenic switch in tumors*. Oncogene 2006, 25(44): 5969-84.
- 9. Jeon, J., Lee, H., Park, H. et al. *Phosphorylation of focal adhesion kinase at tyrosine 407 negatively regulates Ras transformation of fibroblasts*. Biochem Biophys Res Commun 2007, 364(4): 1062-6.
- Lim, Y., Park, H., Jeon, J. et al. Focal adhesion kinase is negatively regulated by phosphorylation at tyrosine 407. J Biol Chem 2007, 282(14): 10398-404.
- 11. Ma, A., Richardson, A., Schaefer, E.M., Parsons, J.T. Serine phosphorylation of focal adhesion kinase in interphase and mitosis: a possible role in modulating binding to p130(Cas). Mol Biol Cell 2001, 12(1): 1-12.
- 12. Parsons, J.T. Focal adhesion kinase: The first ten years. J Cell Sci 2003, 116(Pt. 8): 1409-16.
- Park, A.Y.J., Shen, T.-L., Chien, S., Guan, J.-L. Role of focal adhesion kinase Ser-732 phosphorylation in centrosome function during mitosis. J Biol Chem 2009, 284(14): 9418-25.

- Jacamo, R., Jiang, X., Lunn, J.A., Rozengurt, E. FAK phosphorylation at Ser-843 inhibits Tyr-397 phosphorylation, cell spreading and migration. J Cell Physiol 2007, 210(2): 436-44.
- van Nimwegen, M.J., Huigsloot, M., Camier, A., Tijdens, I.B., van de Water, B. Focal adhesion kinase and protein kinase B cooperate to suppress doxorubicin-induced apoptosis of breast tumor cells. Mol Pharmacol 2006, 70(4): 1330-9.
- 16. Zhao, J.H., Reiske, H., Guan, J.L. Regulation of the cell cycle by focal adhesion kinase. J Cell Biol 1998, 143(7): 1997-2008.
- Zhao, J., Pestell, R., Guan, J.L. Transcriptional activation of cyclin D1 promoter by FAK contributes to cell cycle progression. Mol Biol Cell 2001, 12(12): 4066-77.
- Yamamoto, D., Sonoda, Y., Hasegawa, M., Funakoshi-Tago, M., Aizu-Yokota, E., Kasahara, T. FAK overexpression upregulates cyclin D3 and enhances cell proliferation via the PKC and Pl3-kinase-Akt pathways. Cell Signal 2003, 15(6): 575-83.
- Lim, S.-T., Mikolon, D., Stupack, D.G., Schlaepfer, D.D. FERM control of FAK function: Implications for cancer therapy. Cell Cycle 2008, 7(15): 2306-14.
- 20. Golubovskaya, V.M., Cance, W.G. Focal adhesion kinase and p53 signaling in cancer cells. Int Rev Cytol 2007, 263: 103-53.
- 21. Lim, S.-T., Chen, X.L., Lim, Y. et al. *Nuclear FAK promotes cell proliferation and survival through FERM-enhanced p53 degradation*. Mol Cell 2008, 29(1): 9-22.
- 22. Schlaepfer, D.D., Mitra, S.K. Multiple connections link FAK to cell motility and invasion. Curr Opin Genet Dev 2004, 14(1): 92-101.
- 23. Wozniak, M.A., Modzelewska, K., Kwong, L., Keely, P.J. Focal adhesion regulation of cell behavior. Biochim Biophys Acta 2004, 1692(2-3): 103-19.
- 24. Xu, Y., Benlimame, N., Su, J., He, Q., Alaoui-Jamali, M.A. Regulation of focal adhesion turnover by ErbB signalling in invasive breast cancer cells. Br J Cancer 2009, 100(4): 633-43.
- 25. Chan, K.T., Cortesio, C.L., Huttenlocher, A. FAK alters invadopodia and focal adhesion composition and dynamics to regulate breast cancer invasion. J Cell Biol 2009, 185(2): 357-70.
- Papusheva, E., Mello de Queiroz, F., Dalous, J. et al. Dynamic conformational changes in the FERM domain of FAK are involved in focal-adhesion behavior during cell spreading and motility. J Cell Sci 2009, 122(Pt. 5): 656-66.
- Cohen, L.A., Guan, J.-L. Residues within the first subdomain of the FERM-like domain in focal adhesion kinase are important in its regulation. J Biol Chem 2005, 280(9): 8197-207.
- Chen, S.-Y., Chen, H.-C. Direct interaction of focal adhesion kinase (FAK) with Met is required for FAK to promote hepatocyte growth factor-induced cell invasion. Mol Cell Biol 2006, 26(13): 5155-67.
- 29. Hauck, C.R., Sieg, D.J., Hsia, D.A. et al. Inhibition of focal adhesion kinase expression or activity disrupts epidermal growth factor-stimulated signaling promoting the migration of invasive human carcinoma cells. Cancer Res 2001, 61(19): 7079-90.
- Hauck, C.R., Hsia, D.A., Puente, X.S., Cheresh, D.A., Schlaepfer, D.D. FRNK blocks v-Src-stimulated invasion and experimental metastases without effects on cell motility or growth. EMBO J 2002, 21(23): 6289-302.
- Kornberg, L., Fleigel, J. The effects of inducible overexpression of FAKrelated non-kinase (FRNK) on a transformed epithelial cell line. Anticancer Res 2003, 23(1A): 91-7.
- Canel, M., Secades, P., Garzon-Arango, M. et al. Involvement of focal adhesion kinase in cellular invasion of head and neck squamous cell carcinomas via regulation of MMP-2 expression. Br J Cancer 2008, 98(7): 1274-84.

- 33. Jones, G., Machado, J. Jr., Tolnay, M., Merlo, A. PTEN-independent induction of caspase-mediated cell death and reduced invasion by the focal adhesion targeting domain (FAT) in human astrocytic brain tumors which highly express focal adhesion kinase (FAK). Cancer Res 2001, 61(15): 5688-91.
- 34. Johnson, T.R., Khandrika, L., Kumar, B. et al. *Focal adhesion kinase controls aggressive phenotype of androgen-independent prostate cancer*. Mol Cancer Res 2008, 6(10): 1639-48.
- 35. Owens, L.V., Xu, L., Craven, R.J. et al. *Overexpression of the focal adhesion kinase (p125FAK) in invasive human tumors*. Cancer Res 1995, 55(13): 2752-5.
- 36. Golubovskaya, V.M., Kweh, F.A., Cance, W.G. Focal adhesion kinase and cancer. Histol Histopathol 2009, 24(4): 503-10.
- 37. Zhao, J., Guan, J.-L. Signal transduction by focal adhesion kinase in cancer. Cancer Metastasis Rev 2009, 28(1-2): 35-49.
- 38. Oktay, M.H., Oktay, K., Hamele-Bena, D., Buyuk, A., Koss, L.G. Focal adhesion kinase as a marker of malignant phenotype in breast and cervical carcinomas. Hum Pathol 2003, 34(3): 240-5.
- 39. Behmoaram, E., Bijian, K., Jie, S. et al. Focal adhesion kinase-related proline-rich tyrosine kinase 2 and focal adhesion kinase are co-overexpressed in early-stage and invasive ErbB-2-positive breast cancer and cooperate for breast cancer cell tumorigenesis and invasiveness. Am J Pathol 2008, 173(5): 1540-50.
- Cance, W.G., Harris, J.E., Iacocca, M.V. et al. Immunohistochemical analyses of focal adhesion kinase expression in benign and malignant human breast and colon tissues: Correlation with preinvasive and invasive phenotypes. Clin Cancer Res 2000, 6(6): 2417-23.
- 41. Garcia, S., Dales, J.-P., Charafe-Jauffret, E. et al. Overexpression of c-Met and of the transducers PI3K, FAK and JAK in breast carcinomas correlates with shorter survival and neoangiogenesis. Int J Oncol 2007, 31(1): 49-58.
- 42. Madan, R., Smolkin, M.B., Cocker, R., Fayyad, R., Oktay, M.H. Focal adhesion proteins as markers of malignant transformation and prognostic indicators in breast carcinoma. Hum Pathol 2006, 37(1): 9-15.
- 43. Lark, A.L., Livasy, C.A., Calvo, B. et al. Overexpression of focal adhesion kinase in primary colorectal carcinomas and colorectal liver metastases: Immunohistochemistry and real-time PCR analyses. Clin Cancer Res 2003, 9(1): 215-22.
- 44. Han, N.M., Fleming, R.Y., Curley, S.A., Gallick, G.E. Overexpression of focal adhesion kinase (p125FAK) in human colorectal carcinoma liver metastases: Independence from c-src or c-yes activation. Ann Surg Oncol 1997, 4(3): 264-8.
- 45. Owens, L.V., Xu, L., Dent, G.A. et al. *Focal adhesion kinase as a marker of invasive potential in differentiated human thyroid cancer*. Ann Surg Oncol 1996, 3(1): 100-5.
- 46. Rovin, J.D., Frierson, H.F. Jr., Ledinh, W., Parsons, J.T., Adams, R.B. Expression of focal adhesion kinase in normal and pathologic human prostate tissues. Prostate 2002, 53(2): 124-32.
- 47. Fujii, T., Koshikawa, K., Nomoto, S. et al. *Focal adhesion kinase is overex*pressed in hepatocellular carcinoma and can be served as an independent prognostic factor. J Hepatol 2004, 41(1): 104-11.
- 48. Itoh, S., Maeda, T., Shimada, M. et al. *Role of expression of focal adhesion kinase in progression of hepatocellular carcinoma*. Clin Cancer Res 2004, 10(8): 2812-7.
- 49. Hess, A.R., Postovit, L.-M., Margaryan, N.V. et al. *Focal adhesion kinase promotes the aggressive melanoma phenotype*. Cancer Res 2005, 65(21): 9851-60.
- Sood, A.K., Coffin, J.E., Schneider, G.B. et al. Biological significance of focal adhesion kinase in ovarian cancer: Role in migration and invasion. Am J Pathol 2004, 165(4): 1087-95.

- 51. Agochiya, M., Brunton, V.G., Owens, D.W. et al. *Increased dosage and amplification of the focal adhesion kinase gene in human cancer cells*. Oncogene 1999, 18(41): 5646-53.
- 52. Gabarra-Niecko, V., Schaller, M.D., Dunty, J.M. FAK regulates biological processes important for the pathogenesis of cancer. Cancer Metastasis Rev 2003, 22(4): 359-74.
- 53. Weiner, T.M., Liu, E.T., Craven, R.J., Cance, W.G. Expression of focal adhesion kinase gene and invasive cancer. Lancet 1993, 342(8878): 1024-5.
- 54. Lark, A.L., Livasy, C.A., Dressler, L. et al. High focal adhesion kinase expression in invasive breast carcinomas is associated with an aggressive phenotype. Mod Pathol 2005, 18(10): 1289-94.
- Peng, X., Wu, X., Druso, J.E. et al. Cardiac developmental defects and eccentric right ventricular hypertrophy in cardiomyocyte focal adhesion kinase (FAK) conditional knockout mice. Proc Natl Acad Sci USA 2008, 105(18): 6638-43.
- McLean, G.W., Komiyama, N.H., Serrels, B. et al. Specific deletion of focal adhesion kinase suppresses tumor formation and blocks malignant progression. Genes Dev 2004, 18(24): 2998-3003.
- 57. Weis, S.M., Lim, S.-T., Lutu-Fuga, K.M. et al. Compensatory role for Pyk2 during angiogenesis in adult mice lacking endothelial cell FAK. J Cell Biol 2008, 181(1): 43-50.
- Lahlou, H., Sanguin-Gendreau, V., Zuo, D. et al. Mammary epithelial-specific disruption of the focal adhesion kinase blocks mammary tumor progression. Proc Natl Acad Sci USA 2007, 104(51): 20302-7.
- 59. Nagy, T., Wei, H., Shen, T.-L. et al. Mammary epithelial-specific deletion of the focal adhesion kinase gene leads to severe lobulo-alveolar hypoplasia and secretory immaturity of the murine mammary gland. J Biol Chem 2007, 282(43): 31766-76.
- 60. Provenzano, P.P., Inman, D.R., Eliceiri, K.W., Beggs, H.E., Keely, P.J. Mammary epithelial-specific disruption of focal adhesion kinase retards tumor formation and metastasis in a transgenic mouse model of human breast cancer. Am J Pathol 2008, 173(5): 1551-65.
- Luo, M., Fan, H., Nagy, T. et al. Mammary epithelial-specific ablation of the focal adhesion kinase suppresses mammary tumorigenesis by affecting mammary cancer stem/progenitor cells. Cancer Res 2009, 69(2): 466-74.
- 62. Abbi, S., Ueda, H., Zheng, C. et al. *Regulation of focal adhesion kinase by a novel protein inhibitor FIP200*. Mol Biol Cell 2002, 13(9): 3178-91.
- 63. Ueda, H., Abbi, S., Zheng, C., Guan, J.L. Suppression of Pyk2 kinase and cellular activities by FIP200. J Cell Biol 2000, 149(2): 423-30.
- 64. Melkoumian, Z.K., Peng, X., Gan, B., Wu, X., Guan, J.-L. Mechanism of cell cycle regulation by FIP200 in human breast cancer cells. Cancer Res 2005, 65(15): 6676-84.
- 65. Chano, T., Kontani, K., Teramoto, K., Okabe, H., Ikegawa, S. *Truncating mutations of RBICC1 in human breast cancer*. Nat Genet 2002, 31(3): 285-8.
- 66. Wang, D., Grammer, J.R., Cobbs, C.S. et al. *p125 focal adhesion kinase promotes malignant astrocytoma cell proliferation in vivo.* J Cell Sci 2000, 113(Pt. 23): 4221-30.
- 67. Abdel-Ghany, M., Cheng, H.-C., Elble, R.C., Pauli, B.U. Focal adhesion kinase activated by beta(4) integrin ligation to mCLCA1 mediates early metastatic growth. J Biol Chem 2002, 277(37): 34391-400.
- van Nimwegen, M.J., Verkoeijen, S., van Buren, L., Burg, D., van de Water,
 Requirement for focal adhesion kinase in the early phase of mammary adenocarcinoma lung metastasis formation. Cancer Res 2005, 65(11): 4698-706.
- 69. Mitra, S.K., Schlaepfer, D.D. *Integrin-regulated FAK-Src signaling in normal and cancer cells*. Curr Opin Cell Biol 2006, 18(5): 516-23.
- 70. Mitra, S.K., Lim, S.T., Chi, A., Schlaepfer, D.D. Intrinsic focal adhesion kinase activity controls orthotopic breast carcinoma metastasis via the reg-

- ulation of urokinase plasminogen activator expression in a syngeneic tumor model. Oncogene 2006, 25(32): 4429-40.
- 71. Benlimame, N., He, Q., Jie, S. et al. *FAK signaling is critical for ErbB-2/ErbB-3 receptor cooperation for oncogenic transformation and invasion.* J Cell Biol 2005, 171(3): 505-16.
- 72. Pylayeva, Y., Gillen, K.M., Gerald, W., Beggs, H.E., Reichardt, L.F., Giancotti, F.G. Ras- and PI3K-dependent breast tumorigenesis in mice and humans requires focal adhesion kinase signaling. J Clin Invest 2009, 119(2): 252-66.
- 73. Han, E.K.-H., McGonigal, T., Wang, J., Giranda, V.L., Luo, Y. Functional analysis of focal adhesion kinase (FAK) reduction by small inhibitory RNAs. Anticancer Res 2004, 24(6): 3899-905.
- 74. Jiang, X., Jacamo, R., Zhukova, E., Sinnett-Smith, J., Rozengurt, E. RNA interference reveals a differential role of FAK and Pyk2 in cell migration, leading edge formation and increase in focal adhesions induced by LPA in intestinal epithelial cells. J Cell Physiol 2006, 207(3): 816-28.
- 75. Tsutsumi, K., Kasaoka, T., Park, H.-M., Nishiyama, H., Nakajima, M., Honda, T. *Tumor growth inhibition by synthetic and expressed siRNA targeting focal adhesion kinase*. Int J Oncol 2008, 33(1): 215-24.
- 76. Lipinski, C.A., Tran, N.L., Viso, C. et al. Extended survival of Pyk2 or FAK deficient orthotopic glioma xenografts. J Neurooncol 2008, 90(2): 181-9.
- 77. Li, S., Dong, W., Zong, Y. et al. *Polyethylenimine-complexed plasmid particles targeting focal adhesion kinase function as melanoma tumor therapeutics*. Mol Ther 2007, 15(3): 515-23.
- 78. Halder, J., Kamat, A.A., Landen, C.N. Jr. et al. Focal adhesion kinase targeting using in vivo short interfering RNA delivery in neutral liposomes for ovarian carcinoma therapy. Clin Cancer Res 2006, 12(16): 4916-24.
- 79. Shintani, S., Li, C., Mihara, M., Nakashiro, K.-I., Hamakawa, H. *Gefitinib* ('Iressa'), an epidermal growth factor receptor tyrosine kinase inhibitor, mediates the inhibition of lymph node metastasis in oral cancer cells. Cancer Lett 2003, 201(2): 149-55.
- 80. Liu, Y., Kyle, E., Lieberman, R., Crowell, J., Kellof, G., Bergan, R.C. Focal adhesion kinase (FAK) phosphorylation is not required for genistein-induced FAK-beta-1-integrin complex formation. Clin Exp Metastasis 2000. 18(3): 203-12.
- 81. Matsumoto, K., Nakamura, T., Kramer, R.H. Hepatocyte growth factor/scatter factor induces tyrosine phosphorylation of focal adhesion kinase (p125FAK) and promotes migration and invasion by oral squamous cell carcinoma cells. J Biol Chem 1994, 269(50): 31807-13.
- 82. Vultur, A., Buettner, R., Kowolik, C. et al. SKI-606 (bosutinib), a novel Src kinase inhibitor, suppresses migration and invasion of human breast cancer cells. Mol Cancer Ther 2008, 7(5): 1185-94.
- 83. Johnson, F.M., Saigal, B., Talpaz, M., Donato, N.J. Dasatinib (BMS-354825) tyrosine kinase inhibitor suppresses invasion and induces cell cycle arrest and apoptosis of head and neck squamous cell carcinoma and non-small cell lung cancer cells. Clin Cancer Res 2005, 11(19, Pt. 1): 6924-32.
- Shi, Q., Hjelmeland, A.B., Keir, S.T. et al. A novel low-molecular weight inhibitor of focal adhesion kinase, TAE226, inhibits glioma growth. Mol Carcinog 2007, 46(6): 488-96.

- 85. Slack-Davis, J.K., Martin, K.H., Tilghman, R.W. et al. *Cellular characterization of a novel focal adhesion kinase inhibitor*. J Biol Chem 2007, 282(20): 14845-52.
- 86. Roberts, W.G., Ung, E., Whalen, P. et al. *Antitumor activity and pharma-cology of a selective focal adhesion kinase inhibitor, PF-562,271*. Cancer Res 2008, 68(6): 1935-44.
- 87. Bagi, C.M., Roberts, G.W., Andresen, C.J. *Dual focal adhesion kinase/Pyk2 inhibitor has positive effects on bone tumors: Implications for bone metastases.* Cancer 2008, 112(10): 2313-21.
- Siu L.L., Burris H.A., Mileshkin L.R. et al. A phase I clinical, pharmacokinetic (PK) and pharmacodynamic (PD) evaluation of PF-00562271 targeting focal adhesion kinase (FAK) in patients (pts) with advanced solid tumors.
 J Clin Oncol [44th Annu Meet Am Soc Clin Oncol) (ASCO) (May 31-June 3, Chicago) 2008] 2008, 26(15, Suppl.): Abst 3534.
- Halder, J., Lin, Y.G., Merritt, W.M. et al. Therapeutic efficacy of a novel focal adhesion kinase inhibitor TAE226 in ovarian carcinoma. Cancer Res 2007, 67(22): 10976-83.
- 90. Watanabe, N., Takaoka, M., Sakurama, K. et al. *Dual tyrosine kinase inhibitor for focal adhesion kinase and insulin-like growth factor-I receptor exhibits anticancer effect in esophageal adenocarcinoma in vitro and in vivo*. Clin Cancer Res 2008, 14(14): 4631-9.
- 91. Lietha, D., Eck, M.J. Crystal structures of the FAK kinase in complex with TAE226 and related bis-anilino pyrimidine inhibitors reveal a helical DFG conformation. PLoS ONE 2008, 3(11): e3800.
- 92. Sakurama, K., Noma, K., Takaoka, M. et al. *Inhibition of focal adhesion kinase as a potential therapeutic strategy for imatinib-resistant gastrointestinal stromal tumor*. Mol Cancer Ther 2009, 8(1): 127-34.
- 93. Chatzizacharias, N.A., Kouraklis, G.P., Theocharis, S.E. *Focal adhesion kinase: A promising target for anticancer therapy.* Expert Opin Ther Targets 2007, 11(10): 1315-28.
- 94. Han, E.K.-H., McGonigal, T. *Role of focal adhesion kinase in human cancer: A potential target for drug discovery.* Anticancer Agents Med Chem 2007, 7(6): 681-4.
- 95. van Nimwegen, M.J., van de Water, B. *Focal adhesion kinase: A potential target in cancer therapy*. Biochem Pharmacol 2007, 73(5): 597-609.
- 96. Parsons, J.T., Slack-Davis, J., Tilghman, R., Roberts, W.G. Focal adhesion kinase: Targeting adhesion signaling pathways for therapeutic intervention. Clin Cancer Res 2008, 14(3): 627-32.
- 97. Li, S., Hua, Z.-C. FAK expression regulation and therapeutic potential. Adv Cancer Res 2008, 101: 45-61.
- Golubovskaya, V.M., Virnig, C., Cance, W.G. TAE226-induced apoptosis in breast cancer cells with overexpressed Src or EGFR. Mol Carcinog 2008, 47(3): 222-34.
- 99. Golubovskaya, V.M., Nyberg, C., Zheng, M. et al. A small molecule inhibitor, 1,2,4,5-benzenetetraamine tetrahydrochloride, targeting the y397 site of focal adhesion kinase decreases tumor growth. J Med Chem 2008, 51(23): 7405-16.